

Benthic Studies in Alice Arm and Hastings Arm, B.C. in Relation to Mine Tailings Dispersal

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by

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ABSTRACT

Kathman, R.D., R.O. Brinkhurst, R.E. Woods, and D.C. Jeffries. 1983. Benthic studies in Alice Arm and Hastings Arm, B.C. in relation to mine tailings dispersal. Can. Tech. Rep. Hydrogr. Ocean Sci. 22, 1-30.

Identification of benthic invertebrates was performed on samples collected in Alice Arm and Hastings Arm, British Columbia, in October 1982 to determine distribution of settled mine tailings and the effect on the benthos. Taxonomic and hierarchical analyses of these organisms indicated that mine tailings were affecting the benthic composition and density in the middle to lower section (towards the sill) of Alice Arm. Sediment data from other sources support these findings.

Key words: benthic invertebrates, mine tailings, sedimentation, fjords.

RÉSUMÉ

Kathman, R.D., R.O. Brinkhurst, R.E. Woods, and D.C. Jeffries. 1983. Benthic studies in Alice Arm and Hastings Arm, B.C. in relation to mine tailings dispersal. Can. Tech. Rep. Hydrogr. Ocean Sci. 22, 1-30.

L'identification d'invertébrés benthiques a été faite à partir d'échantillons recueillis dans le bras Alice et le bras Hastings (Colombie-Britannique) en octobre 1982 pour déterminer la distribution de stériles miniers déposés et l'effet sur le benthos. L'analyse taxonomique de ces organismes et de leur hiérarchie montre que les stériles miniers ont une incidence sur la composition et la densité du benthos à partir du milieu jusqu'à la partie inférieure (vers le seuil) du bras Alice. Les données sur les sédiments provenant d'autres sources appuient ces conclusions.

Mots-cles: invertébrés benthiques, stériles miniers, sédimentation, fjords.

ACKNOWLEDGEMENTS

E.V.S. Consultants would like to thank all the people who contributed to this project. Verifications of organisms were done by the following: Dr. Craig Staude - Amphipoda; Dr. Frank Bernard - Mollusca; Mr. Eugene Ruff - Polychaeta; Mr. Steve Stricker - Nemertea; and Dr. William Austin - Echinodermata and minor phyla. Mr. Jean Coustalin performed the initial taxonomic identifications. Several scientists provided additional information; we would especially like to thank Drs. Jeff Thompson and Rob Macdonald, and Mr. Darcy Goyette for their input. E.V.S. Consultants personnel included Dr. Peter Chapman, who reviewed the manuscript and added valuable comments; Mss. Marla Mees and Sarah Irwin, who prepared the report; Ms. Connie Priestley, who assisted with data and draft report preparation; and Mr. Paul Wheeler, who deserves special thanks for coordination and production of the report. Mss. Deedee Kathman and Roberta Woods were the Principal Investigators. Cluster analysis was done by Mr. Duncan Jeffries of Quantum Research. Dr. Ralph Brinkhurst assisted with report preparation.

SUMMARY

Benthic invertebrate samples were collected during October 1982 in Alice Arm and Hastings Arm, British Columbia. Composition, density and distribution of the benthos were analyzed in relation to the distribution of settled mine tailings in Alice Arm as determined by corresponding studies.

Transects CC and DD in Alice Arm had very few species, indicating severe stress at these locations, particularly in the north and middle stations at CC and the middle station at DD. Transect EE had more species, although the deep middle station along this transect had reduced numbers, comparable to Transect DD. This indicates that mine tailings are probably beginning to affect the lower section of Alice Arm, and have progressed to the deep middle section of EE. Independent sedimentation data and sediment analyses from other sources agree with these findings.

North and south stations on Transect EE had similar numbers of species and individuals as the stations in Hastings Arm. However, the qualitative taxonomic composition of the fauna was quite different. Greater varieties and abundances of nemertean, polychaetes, gastropods, scaphopods, bivalves and echinoderms occurred in Hastings Arm, while nematodes, amphipods and holothuroids were more diverse and abundant at the north and south stations at EE. These differences were probably not related to the effects of tailings but rather to differences in physical, chemical, and biological factors between the two fjords.

Station FF nearest the sill in Alice Arm was dissimilar in composition and density from all other stations sampled and could not be used for comparisons. A heavy sediment load from Hans Creek which settled at Station FF, as well as large amounts of wood fibers in the substrate, contributed to the uniqueness of benthic invertebrates at this station.

INTRODUCTION

The study of the effect of mine tailings deposition on marine benthic invertebrates in Alice Arm was carried out to indicate the extent and degree of dispersion of tailings along the deeper part of Alice Arm. This study formed one component of an overall study being conducted by the Canadian federal government into the effects of Kitsault mine tailings deposition on the Alice Arm aquatic ecosystem. Other concurrent surveys include zooplankton composition and distribution, sedimentation rates, trace metal concentrations in sediments, and hydrographical data collections.

Tailings discharge into Alice Arm began in April 1981 and ceased with the temporary shut-down of the mill in November 1982. During that period approximately four million tons of process effluent were discharged into Alice Arm (R. Hinder, pers. comm.). A variety of fauna may be affected by these tailings due to toxicity of heavy metal concentrations, increased turbidity limiting light penetration and primary production, and smothering of infaunal organisms.

Samples were collected from a series of transects in Alice Arm at increasing distances from the discharge site, and from adjacent Hastings Arm, an area unaffected by tailings discharge. Benthic invertebrates were sorted from the sediment and the infauna identified. The benthic infaunal data were used to assess possible effects, including extent and severity, of tailings deposition on benthic communities.

TERMS OF REFERENCE

The overall objective of this study was to determine composition, density and distribution of the benthic invertebrate fauna in relation to mine tailings distribution in Alice Arm, British Columbia. Specific objectives were:

1. Sort and identify benthic macroinvertebrates retained by a 1.0 mm mesh screen in samples collected from Alice Arm and Hastings Arm, British Columbia.
2. Count benthic invertebrates retained by a 0.25 mm mesh screen in samples collected from Alice Arm and Hastings Arm, British Columbia.
3. Verify all identified material from 1. above.
4. Perform statistical analyses on identification data from 1., allowing comparisons among stations and between the two basins.
5. Prepare a data report incorporating the above and other pertinent information with data on the distribution of infaunal benthic invertebrates in relation to settled mine tailings in Alice Arm.

METHODS

SAMPLING

Sampling in Alice Arm and Hastings Arm, British Columbia (Fig. 1), was conducted by Dobrocky Seatech Ltd. in October 1982. Two replicates were taken at each station with the exception of Station EEM on predetermined transect lines (Fig. 2), using a 0.1 m² Smith-McIntyre grab. Each sample was sieved through a 1.0 mm screen and preserved with seven percent formalin with phloxine B, an histological stain used to facilitate sorting. Residues from each 1.0 mm sieving were then screened at 0.25 mm and the contents preserved as previously described. Samples were transferred to the E.V.S. Consultants Taxonomy Centre on 12 and 18 November. Pertinent information recorded during the field survey is presented in Appendix A.

SORTING AND QUALITY CONTROL

Upon receipt of the above samples by the E.V.S. Consultants Taxonomy Centre, each 1.0 mm-sieved sample was washed through a 1.0 mm sieve to remove formalin. Aliquots of each sample were examined using a Wild M3 or M5A stereomicroscope until the entire sample was sorted and all organisms removed. Organisms were counted and placed into separate containers filled with alcohol according to the following major taxonomic groups: Amphipoda, Other Crustacea, Polychaeta, Oligochaeta, Mollusca, Nematoda, and Others. Pertinent information for each sample was recorded using the format shown in Appendix B. Sample residues were represerved with seven percent formalin.

Each 0.25 mm-sieved sample was washed through a 0.25 mm sieve to remove formalin and examined as above. Organisms were counted according to major taxonomic groups (for example, Foraminifera, Nematoda, Polychaeta, Ostracoda, Harpacticoida). Only the Oligochaeta were removed, preserved in ethanol and given to the Scientific Authority. A separate information sheet was compiled for each sample using the format shown in Appendix C. Residues were re-preserved with seven percent formalin for possible future analyses.

To ensure quality control of sorting, thirty-two percent (10 samples) of the 1.0 mm residue samples were independently resorted following the procedure outlined above. If additional organisms were found, they were placed in the appropriately labelled vial and information on the sorting sheets was changed accordingly. A separate sheet for each quality control check was completed, in the format shown in Appendix D.

Quality control data for sorting of 1.0 mm screened samples are provided in Table 1. Percent error ranged from 0 to 4.2, well within the requisite limits for accuracy of sorting benthic invertebrate samples.

IDENTIFICATION AND VERIFICATION

All organisms were identified to the lowest possible taxonomic level consistent with the presently-available literature. Identifications were accomplished using either a Wild M5A stereomicroscope or a Leitz Laborlux compound microscope. A list of the taxonomic references used for identifications and verifications is provided.

Fig. 1. Map showing general location of study area.



Fig. 2. Station locations in Alice Arm and Hastings Arm, British Columbia.

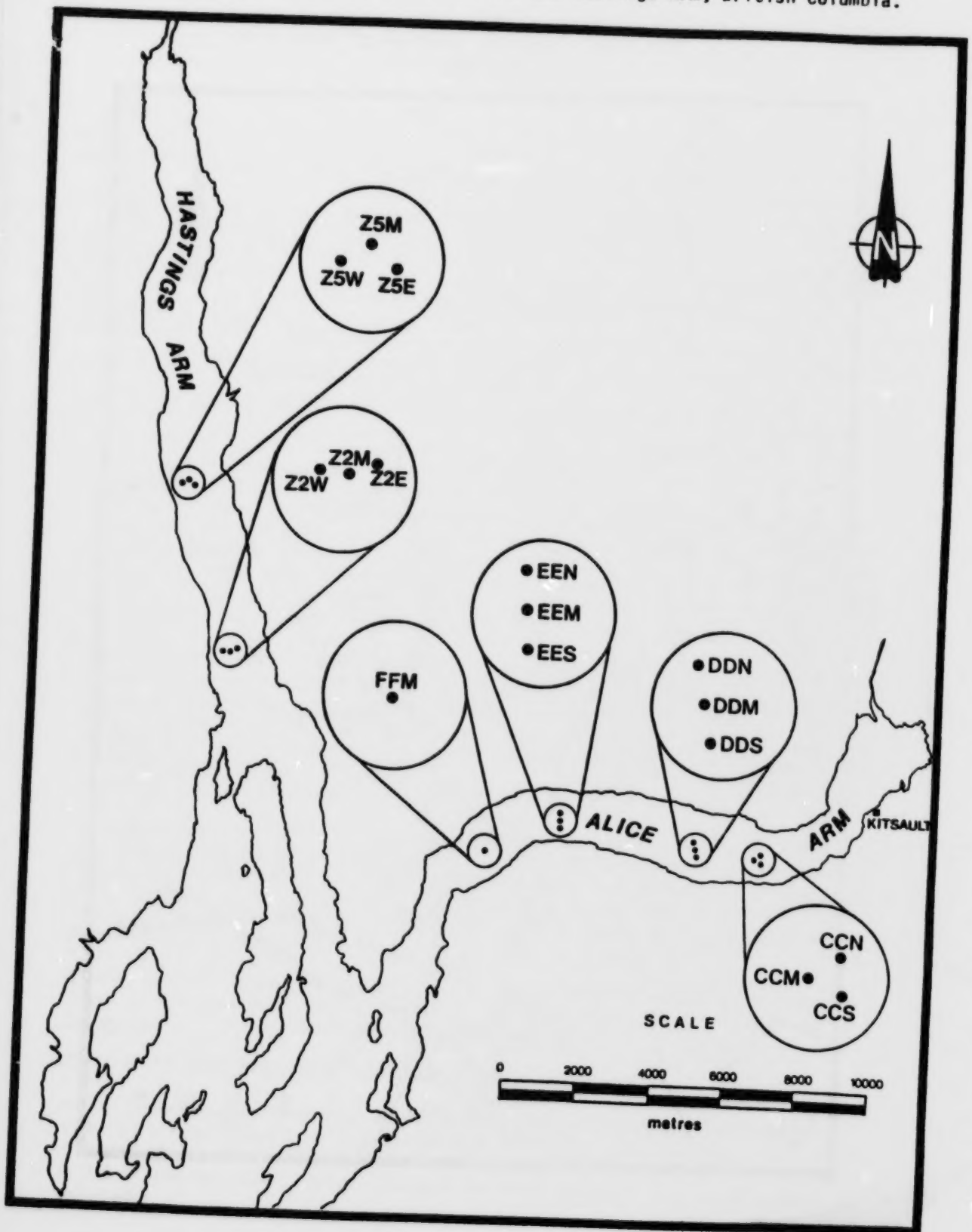


TABLE 1. Quality control (QC) for sorting benthic invertebrates from sediments sieved through a 1.0 mm screen.

Sample identification	Total number of individuals during sorting	Total number of individuals after QC	Percent error for sample
		2	0
CCN-A	2	5	0
CCM-B	5	77	3.9
CCS-A	74	59	1.7
DDN-B	57	3	0
DDM-A	3	333	0.3
EEN-A	332	104	0
EEM-A	104	119	4.2
Z2W-B	114	101	1.0
Z5E-A	100	210	1.0
Z5W-B	208		

Representative samples of organisms identified to the generic or specific level were sent to recognized taxonomic experts (approved by the Scientific Authority) for verification. If the original identification was incorrect, all samples of that genus or species were re-examined to ensure accuracy. In addition, all samples identified to the familial level were sent to experts for possible identification to a lower taxonomic unit. All necessary changes were incorporated into the original identification sheets and vial labels.

A representative sample of each genus/species was separated from the entire collection and presented to the Scientific Authority as a reference set for future use. The identity of all reference material was further verified by appropriate taxonomic experts.

HIERARCHICAL ANALYSIS

Two-way cluster (hierarchical) analysis was performed on the data using the cluster analysis program "CGROUP", allowing comparison among species and among stations. This program also allowed comparison among species and between Alice Arm and Hastings Arm. The program uses the grouping algorithm as described by Ward (1963) and follows recognized and accepted techniques for analysis of benthic data. Replicate samples for each station were averaged in order to produce an appropriate data base for use in the cluster analysis. This also simplified the problem associated with the single sample for Station EEM. Data were also entered individually and are therefore on file and readily accessible for possible future analyses.

OTHER RELATED STUDIES

Other agencies and individuals were contacted to identify pertinent past and current studies in Alice Arm and in other areas impacted by mine tailings (for example, Rupert Inlet). Respondents included Drs. D. Mackas, R. Macdonald, and J. Thompson and Mr. D. Stucchi (Institute of Ocean Sciences), Drs. J.-McInerney and D. Ellis (University of Victoria), Dr. S. Calvert (University of British Columbia), Dr. J. Patterson (Amax of Canada Ltd.), Mr. D. Goyette (Environmental Protection Service), and Mr. W. Drinnan (Dobrocky Seatech Ltd.). A partial listing of studies and reports is given in Appendix E. However, given the terms of reference for this study, a comprehensive review of all pertinent information was not possible.

RESULTS

BENTHIC INVERTEBRATES - 0.25 mm SIEVE SAMPLES

The results of simple scanning of the invertebrates retained by a 0.25 mm sieve are presented in Appendices F-1 through F-4. They are summarized for all taxa in Table 2. These samples were initially retained to ascertain if differences in benthic distribution and density in relation to tailings dispersal would be observed between these and the 1.0 mm sieved samples. Although the 1.0 mm sieved samples only accounted for approximately 23 percent of the total benthic invertebrates collected in Alice Arm and Hastings Arm, similar patterns between each set of samples among stations and transects can be observed. For example, for both sieve sizes fewer numbers of organisms were collected at the middle stations on Transects DD and EE than at the north and south stations, and the north and middle stations on Transect CC had fewer individuals than the south station.

Nematodes (which are not readily identifiable) comprised 70 percent of the small organisms found in Alice Arm. If the contribution of these organisms to overall numbers in samples of both sieve sizes is discounted, the patterns of similarity become apparent (see Table 2). The data indicate that benthic samples screened with 1.0 mm mesh will provide enough information to accurately assess trends and patterns in benthic composition and distribution.

BENTHIC INVERTEBRATES - 1.0 mm SIEVE SAMPLES

A listing of all benthic invertebrates collected during this study is given in Appendix G. The number of organisms for each sample and the mean number of organisms for each station in Alice Arm and Hastings Arm are presented in Appendices H and I, respectively. The cluster analysis performed on these data clearly differentiates three groups of stations: Group 1 - Alice Arm not including Station FFM; Group 2 - Hastings Arm; and Group 3 - Station FFM in Alice Arm (see Figure 3). These groupings are based on the species, numbers of taxa and numbers of individuals collected. Three subgroups can be differentiated in Group 1. Stations CCN, DDM and CCM are very similar, having a very poor benthic infaunal composition. Visual observations made from Pisces IV and trace metal analyses of sediments in cores verify a more northern circulation of tailings at Transect CC (D. Goyette, pers. comm.). The next subgroup (1.2) comprises the remaining stations in Transects CC and DD which are also quite severely affected by tailings deposition. Stations on Transect EE join stations on CC and DD at Step 6 in the clustering. EEM is the most divergent station on CC and DD at Step 6 in the clustering. EEM is the most divergent station on Transect EE and most similar to stations on Transects CC and DD. Figure 4 illustrates a tentative horizontal display of the cluster analysis, showing the similarity among stations in relation to tailings distribution. All species collected at Station EEM were also found at EEN and EES, although approximately one-third as many individuals and one-half as many taxa occurred at EEM. This substantial decrease is indicated by the dashed lined in Figure 4. Sedimentation data substantiate settled tailings occurring at Transect EE as shown in Figure 5 (R. Macdonald, pers. comm.). Trace metal analyses indicate that surface sediments in 1981 at Station EEM contain much higher concentrations than at either EEN or EES, and are comparable to concentrations at Stations CCM and DDM (Goyette and Christie, 1982).

TABLE 2. Comparison of the results of 0.25 mm and 1.0 mm sieve screening of the same benthic sediment samples. All results are given in numbers per m² and are the mean of two samples (except EEM).

	0.25 mm			1.0 mm		
	No. of Individ. per Stat.	No. of Individ. per Trans.	No. of Individ. per Inlet	No. of Individ. per Stat.	No. of Individ. per Trans.	No. of Individ. per Inlet
CCN	675			20		
CCM	230	3,260		30	715	
CCS	2,355			665		
DDN	3,565			520		
DDM	775	7,235	31,200	25	1,630	9,950
DOS	2,895			1,085		
EEN	11,140			3,125		
EEM	1,755	20,705		1,040	7,605	
EES	7,810			3,440		
FFM*	64,380			12,565		
Z2E	11,725			1,835		
Z2M	6,415	22,455		1,255	5,175	
Z2W	4,315			2,085		
			56,355			10,305
Z5E	14,830			1,310		
Z5M	8,715	33,900		2,075	5,130	
Z5W	10,355			1,745		

*Station FFM considered a separate and distinct area.

Fig. 3. Cluster analysis station groupings.

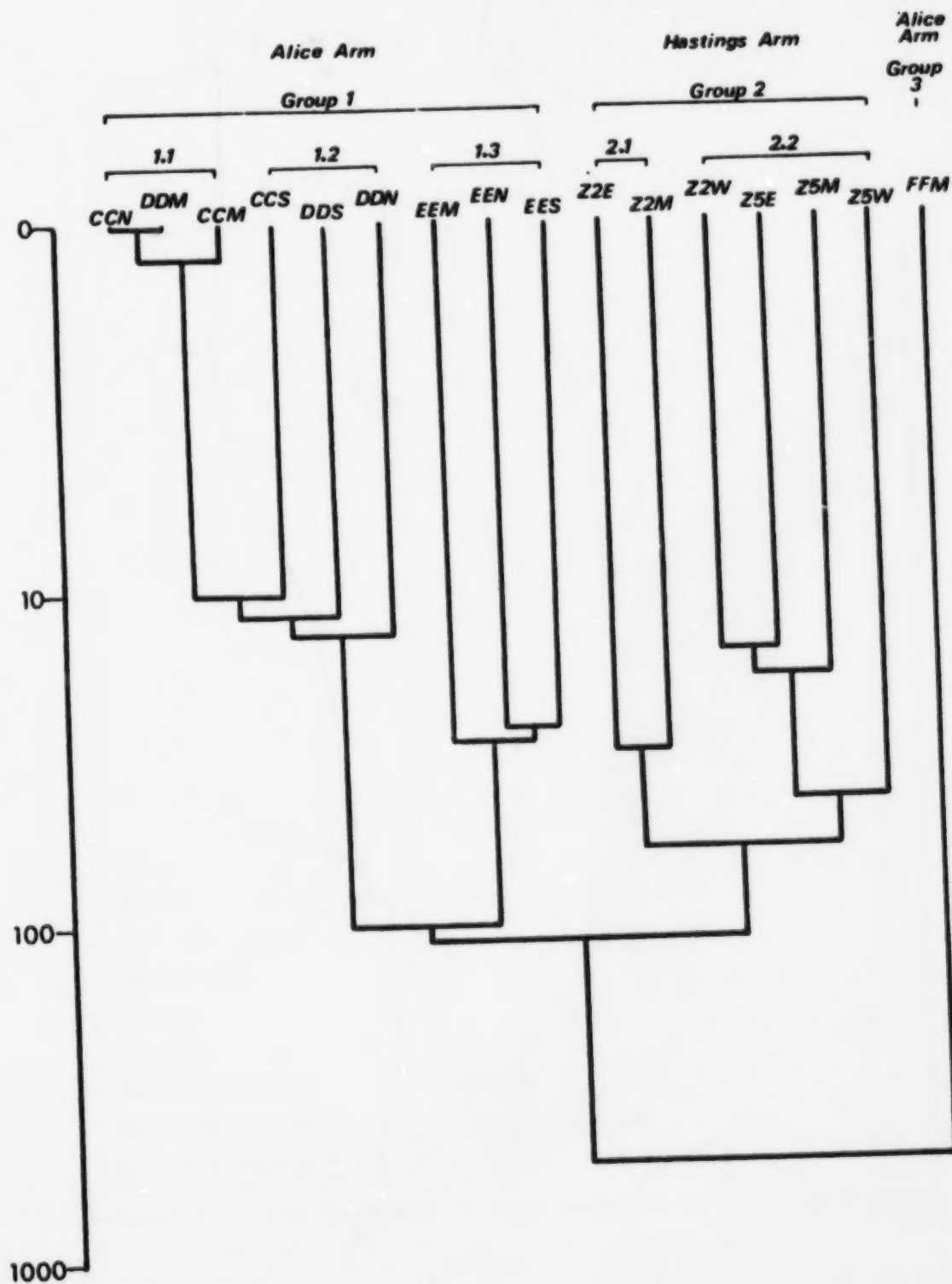


Fig. 4. Tentative horizontal display of cluster analysis of station groupings derived from data in Fig. 3.

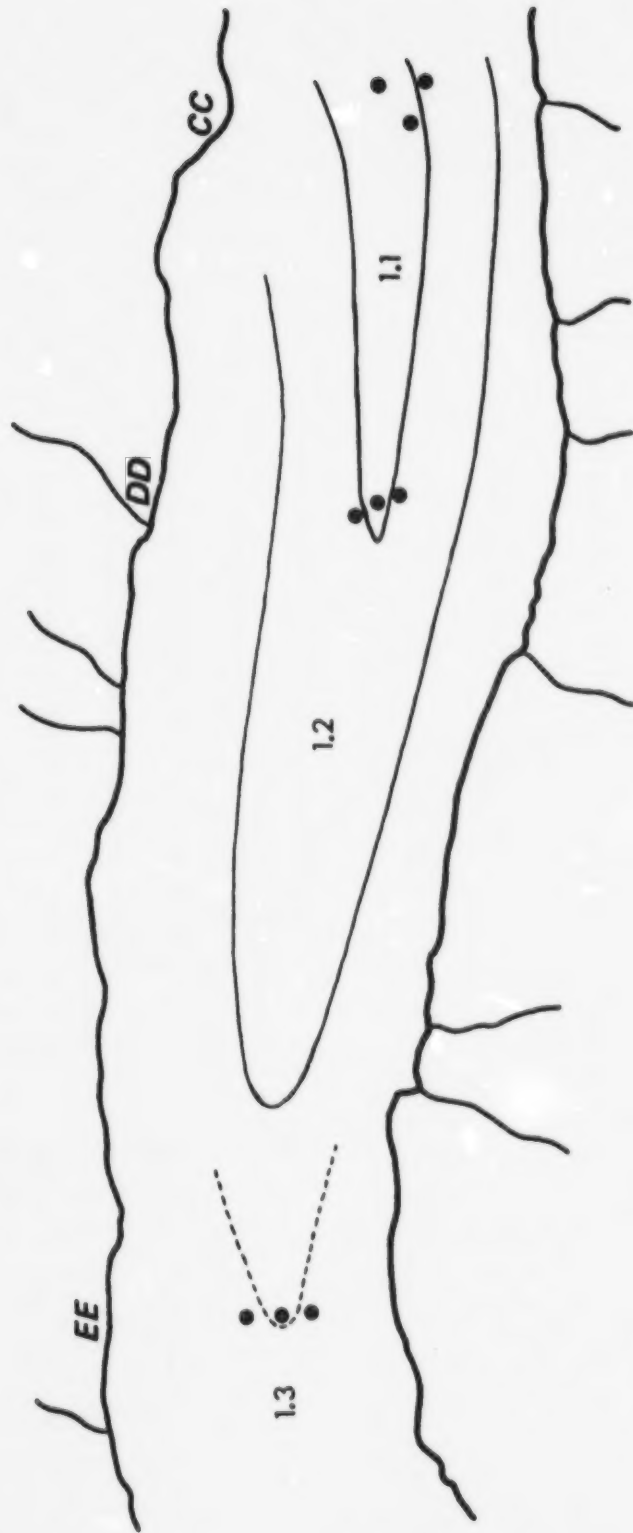
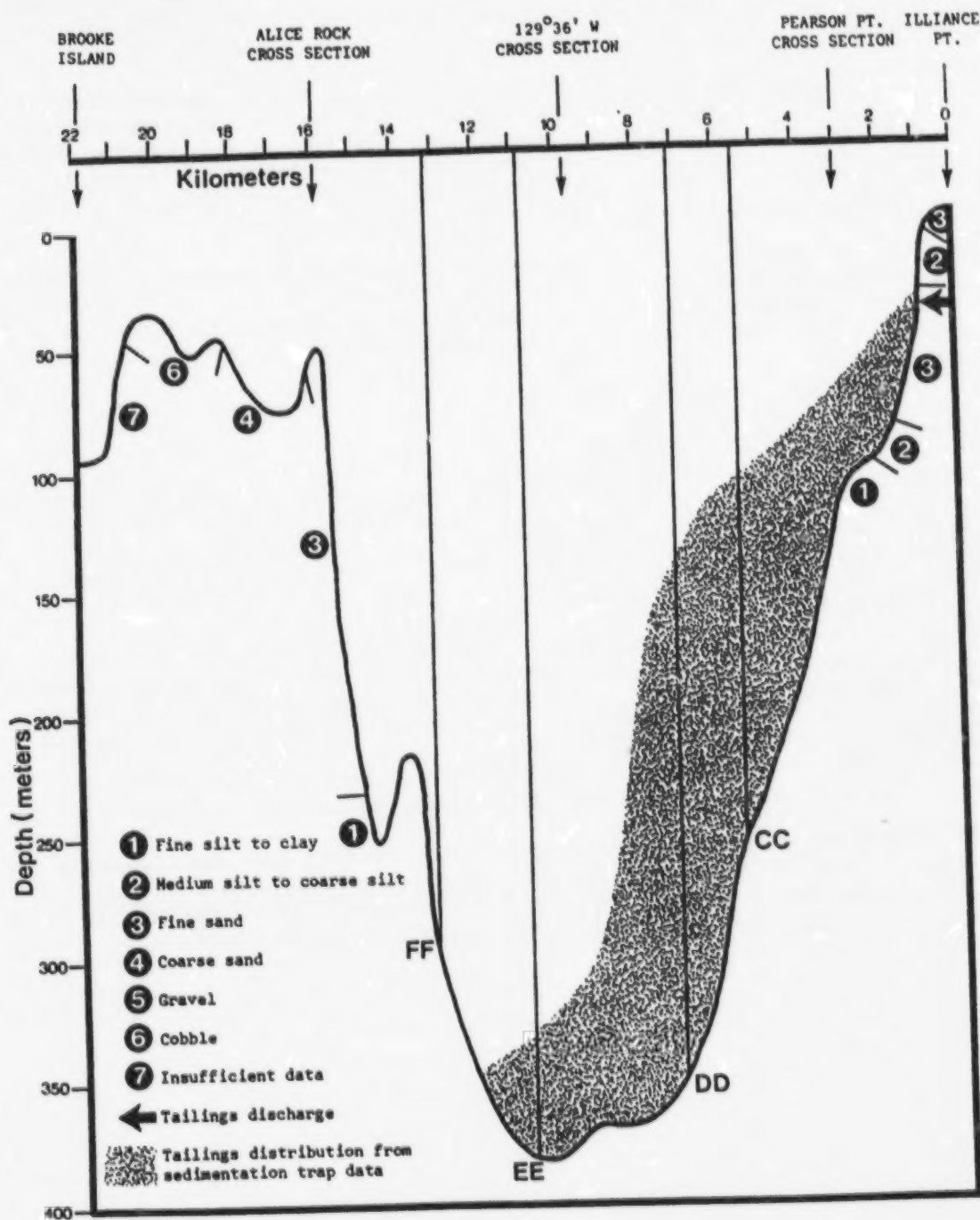


Fig. 5. Longitudinal section of Alice Arm showing generalized sediment distribution, sampling locations, and tailings distribution. Data from R.W. Macdonald and J.L. Littlepage.



While the cluster analysis delineates two subgroups of stations (2.1 and 2.2) within Hastings Arm, we feel the degree of dissimilarity is not a significant factor for comparison with Alice Arm. Group 2 stations are clearly differentiated from Group 1 stations at Step 14. Speciation and density of the benthos as seen in Table 3 and Appendices H and I are also distinct between the two inlets.

Station FFM in Alice Arm forms Group 3, which is widely divergent from all other stations. This station contained higher numbers of individuals and numbers of taxa than all other stations combined in either Alice Arm or Hastings Arm (Table 3). At least twice as many endemic taxa (organisms restricted to a particular station in this study) were found at this station compared to each other transect.

The number of individuals found in the northern and middle samples at CC, and the middle samples at DD and EE in Alice Arm were an order of magnitude lower than the number of individuals found in the equivalent southern samples at CC, and the equivalent northern and southern samples at DD and EE, as shown below.

Transect	<u>CC</u>			<u>DD</u>			<u>EE</u>		
	<u>N</u>	<u>M</u>	<u>S</u>	<u>N</u>	<u>M</u>	<u>S</u>	<u>N</u>	<u>M</u>	<u>S</u>
Number of individuals per m ²	20	30	670	520	25	1,085	3,125	1,040	3,440

Polychaetes were the most abundant organisms at Transects CC, DD and FF, while polychaetes and molluscs were equally abundant at EE in Alice Arm (Table 3). Nephtys cornuta cornuta, Levinsenia gracilis, Spiophanes sp. and Myriochele oculata were the overall dominant polychaetes. Transenella tantilla and Diplodonta orbella were the most abundant bivalves. The fauna at Transects CC and DD, with the exception of five nemertean at Station DDN, consisted of only four major groups: polychaetes, crustaceans, molluscs and echinoderms.

Polychaetes were the most abundant organisms in Hastings Arm, represented by large numbers of N. cornuta cornuta, Aricidea lopezi lopezi, L. gracilis, Chaetozone setosa, Cossura soyeri and Cirratulidae (Table 3). The bivalves T. tantilla, D. orbella and Nucula tenuis, and ophiuroids were also common.

TABLE 3. Summary of benthic invertebrate composition and density for Alice Arm and Hastings Arm.

Mean number of individuals per m ²	Regions				
	Alice Arm			Hastings Arm	Alice Arm
	CC	DD	EE	Z2 + Z5	FF
All individuals	238	543	2,843	1,718	12,565
Polychaeta spp.	216	112	922	1,213	11,170
Mollusca spp.	8	73	1,648	306	335
Crustacea spp.	7	12	86	59	90
Total number of taxa per transect					
All taxa	27	25	52	69	93
Polychaeta spp.	18	12	22	35	61
Mollusca spp.	3	5	15	14	11
Crustacea spp.	3	6	9	12	7
Endemic taxa*	0	4	8	14	29
Endemic Polychaeta spp.	0	0	1	4	16
Endemic Mollusca spp.	0	0	2	4	4
Endemic Crustacea spp.	0	3	4	2	3
No. of samples	6	6	5	12	2

*Species limited to a transect in this study.

DISCUSSION

Visual examination of the data indicates a depressed number and variety of benthic infaunal species at all stations on Transects CC and DD in Alice Arm. Field observations (S. Byers, pers. comm.) suggest that deposits of mine tailings were present in the sample sediments from these stations. Data on the present areal distribution of mine tailings (Fig. 5; R. Macdonald, pers. comm.) indicate that both these transects are affected by settled tailings deposits. Cluster analysis of station groupings (Fig. 3) agrees with grain size distribution and trace metal concentrations (Goyette and Christie, 1982) and light transmission properties (D. Stucchi, pers. comm.). Suppressed benthic fauna at CCN and CCM, but slightly elevated benthic populations at CCS coincide with the initial northerly tailings plume track from Kitsault (D. Goyette, pers. comm. and D. Stucchi, pers. comm.). These tailings appear to be settling in the deep middle stations at Transects DD and EE, as shown in Figure 4 and evidenced by the reduced numbers of species and individuals at DDM and EEM. Equally high trace metal concentrations at CCN, CCM, DDM and EEM (Goyette and Christie, 1982) suggest that tailings have progressed down the inlet at least to EEM, but have not spread to the north or south sides of the channel in this area.

The benthic fauna at Station FFM was distinct from all other stations in both Alice Arm and Hastings Arm. Cluster analysis, visual observation and benthic composition indicate that this station is not representative of other areas in the inlet. This station is directly affected by Hans Creek with highly variable sediment and associated organic loadings. In October 1982, during the period of sample collection, large sediment inputs (three or four times average transport) were collected at this station location in sedimentation traps at subsurface and near-bottom locations (R. Macdonald, pers. comm.). The substrate also differed from that at other Alice Arm stations, with large amounts of wood fiber observed in the sediment during field sampling (S. Byers, pers. comm.). This station is also located close to the sill at the mouth of Alice Arm, which may produce different circulation patterns, making available more niches and more nutrients. Natural variability in this area would be expected to be high. Consequently, this station may not provide useful information related to effects of mine tailings dispersal in Alice Arm.

Faunal composition and density in Hastings Arm appeared to be distinct from that in Alice Arm, as evidenced in the cluster analysis (Fig. 3). The faunal density and diversity is similar between Hastings Arm Transects Z2 and Z5, and Alice Arm Stations EEN and EES, (Table 3; Appendices H and I). (The major difference between EEN and EES compared to Hastings Arm is the large number of the bivalves Tranzenella tantilla and Nucula tenuis found at EEN and EES.) Hastings Arm thus serves as a useful reference area for comparison of "normal" benthic composition, even though it differs in detail from Alice Arm. Although the two fjords are physically distinct and direct comparisons cannot be made between the benthic fauna, Hastings Arm serves as an important reference inlet on which to compare species groupings and diversity.

Changes in infaunal composition and density can be observed between the data from a 1977 benthic study and the 1982 data set. The mean numbers of taxa and mean numbers of individuals at Transect CC in Alice Arm (identified as Station G9.5 in 1977) have decreased substantially in the 1982 samples, as indicated below.

	Transect CC		Percent reduction
	1977	1982	
Mean number of taxa per sample	23	11	52
Mean number of individuals per m ²	483	238	51

This decrease is even more noticeable if only CCN and CCM (1982) data, in which the mean number of taxa was four and the mean number of individuals was 25, are compared with the 1977 data. Station CCS, however, was similar to G9.5 (23 vs. 23 taxa; 665 vs. 483 individuals, respectively), suggesting that the north and middle stations have been dramatically affected by mine tailings.

Further comparisons between the 1977 and 1982 sampling data for Alice Arm are complicated by the fact that station locations differed. For instance, although the 1982 Transect EE was located in the same general area as the 1977 Station F15, the latter was approximately 20 m deeper and located farther from the tailings discharge. However, comparison of the two data sets indicated substantial differences not totally attributable to location. Of 18 species of polychaetes collected in 1977 and 23 species in 1982, only seven were common to both collections. The 1977 samples showed no clear dominance by particular organisms, while the 1982 benthos was dominated by the bivalve Transenella tantilla (38 percent) and the polychaete Nephtys cornuta cornuta (17 percent).

A number of factors in addition to increased mine tailings dispersal could be responsible for the marked differences between 1977 and 1982 collections. Sampling in 1977 took place during June whereas 1982 samples were collected in October and seasonal differences may be a factor. In addition, 1977 samples were not collected in the same locations as 1982 samples. The closest match between the two collection periods is provided by Transect CC (=Station G9.5) with only minor differences in location (latitude varied less than one minute and longitude less than 15 seconds), but the sampling records indicate that 1977 collection depths were greater than those recorded in 1982 by an average of over 24 meters. Other factors such as natural variability may also be implicated.

Overall, many species of polychaetes collected in 1977 were not found in 1982. But the most abundant sedentary polychaete found in 1982, Levinsenia gracilis, was not collected at all in 1977, indicating that both elimination and addition of species have occurred in Alice Arm. Of the 23 species of polychaetes identified in 1977 and the 28 species identified in 1982, only five were common to both sampling periods.

Previous sampling closer to the Alice Arm sill (Station I 17.5 in 1977) indicated that Station FFM was not representative of other areas during the October sampling. Taxa were approximately threefold higher and numbers of individuals tenfold higher in 1982 when the three sample average in 1977 is compared with the two sample average in 1982. Reasons for variabilities have been discussed previously.

Transects Z2 and Z5 in Hastings Arm were sampled in both 1982 and 1977, allowing comparisons to be made among numbers of taxa, numbers of individuals and species. Numbers of taxa and numbers of individuals were lower in 1977 than in 1982 as shown below.

Year	<u>1977</u>		<u>1982</u>	
Transect	<u>Z2</u>	<u>Z5</u>	<u>Z2</u>	<u>Z5</u>
Mean number of taxa per sample	15	15	36	37
Mean number of individuals per m ²	320	227	1725	1710

These changes in faunal composition and density are apparently typical of temporal changes observed in other fjords. For example, numerous studies related to tailings discharge from Island Copper Mine in Rupert Inlet indicate annual shifts in benthic composition and density (D. Ellis, pers. comm.). No individual taxon was clearly dominant in 1977, but polychaetes were dominant at both transects in 1982 (71 percent), followed by molluscs (18 percent). Nephtys cornuta cornuta, L. gracilis, A. lopezi lopezi and Cirratulidae were dominant polychaetes, and T. tantilla and N. tenuis were the dominant bivalve molluscs.

The 1982 data set indicated that benthic infaunal composition and density in Hastings Arm are representative of northern fjords. Undisturbed stations in Alice Arm (EEN, EES) have similar benthic density and diversity as stations in Hastings Arm, making the latter useful as a reference location for comparisons. The data for Alice Arm indicated that mine tailings in Alice Arm are progressing westward down the inlet and settling in the deepest part of the trough.

Elimination of most benthic invertebrates at Transects CC and DD and a large decrease at Station EEM coincides with sedimentation data on tailings distribution and trace metal concentrations of sediments. The present benthic infaunal data set also provides a basis for future comparisons to evaluate possible faunal changes and to relate these changes to substrate disturbances such as tailings sedimentation.

RECOMMENDATIONS

1. An annual or biennial benthic program should be implemented to monitor the density and distribution of benthic invertebrates in relation to settled mine tailings in Alice Arm. This should include stations in both Alice Arm and Hastings Arm.
2. Additional transects should be established between DD and EE, and EE and FF to provide more detailed information on the dispersal of mine tailings and the effect on benthos.
3. Benthic sampling should be done prior to commencement of mill operations after a period of recovery has taken place, followed by sampling a sufficient time after operation to pinpoint and determine specific effects caused by tailings discharge.

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APPENDIX A
AUXILIARY INFORMATION TAKEN
DURING SAMPLE ACQUISITION

<u>Date</u> <u>(October 1982)</u>	<u>Station*</u>	<u>Avg. Depth</u> <u>(m)</u>	<u>North latitude</u> <u>West longitude</u>
23	CCN	264.5	55°26'40" 129°31'42"
26	CCM	277.5	55°26'36" 129°31'53"
26	CCS	256.0	55°26'33" 129°31'44"
25	DDN	349.5	55°26'48" 129°33'37"
26	DDM	351.0	55°26'44" 129°33'35"
26	DDS	356.0	55°26'41" 129°33'31"
25	EEN	370.0	55°27'12" 129°37'00"
25	EEM**	373.0	55°27'06" 129°37'00"
25	EES	369.0	55°27'00" 129°37'00"
26	FFM	303.0	55°26'30" 129°39'03"
24	Z2E	296.0	55°29'19" 129°45'37"
24	Z2M	296.0	55°29'17" 129°45'50"
24	Z2W	296.0	55°29'14" 129°45'57"
24	Z5E	294.0	55°31'41" 129°46'53"
24	Z5M	295.0	55°31'41" 129°47'02"
24	Z5W	295.0	55°31'38" 129°47'12"

* Stations CCN to FFM are located in Alice Arm; Z2E to Z5W are located in Hastings Arm.

** One sample taken at EEM; two samples at all other stations.

APPENDIX B

INFORMATION SHEET USED DURING
SORTING OF 1.0 mm-SIEVED SAMPLES

Sample identification _____

Date of Collection _____ Sieve Size _____

Sorted by _____ Date sorted _____ No. Hours _____

<u>TAXA</u>	<u>NO. OF INDIVIDUALS</u>
-------------	---------------------------

Amphipoda	_____
-----------	-------

Other Crustacea	_____
-----------------	-------

Polychaeta	_____
------------	-------

Oligochaeta	_____
-------------	-------

Molluska	_____
----------	-------

Nematoda	_____
----------	-------

Others	_____
--------	-------

Substrate description (by percent) _____

Observations/comments _____

APPENDIX C

**INFORMATION SHEET USED DURING
SORTING OF 0.25 mm-SIEVED SAMPLES**

Sample identification _____

Date of Collection _____ Sieve Size _____

Sorted by _____ Date sorted _____ No. Hours _____

<u>Phylum</u>	<u>Class/Order</u>	<u>No. of Individuals</u>
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____

Substrate description (by percent) _____

Observations/comments _____

APPENDIX D
INFORMATION SHEET USED DURING
QUALITY CONTROL OF 1.0 mm SAMPLES

Sample identification _____

Date of Collection _____ Sieve Size _____

Sorted by _____ Date sorted _____ No. Hours _____

QC'd by _____ Date QC'd _____ No. Hours _____

<u>Organism</u>	<u>Number of indiv. for taxon found during QC</u>	<u>No. indiv. for taxon originally found</u>	<u>Percent error for taxon</u>	<u>Total no. of individuals in sample</u>	<u>Percent error for sample</u>
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Total number individuals missed _____

Percent error for total sample _____

APPENDIX E
PERTINENT STUDIES AND REPORTS
PARTIAL LISTING

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- Retech Ltd. Sedimentation of mine tailings in a marine environment.*
- Stucchi, D. Analysis of the Crash Program data (1981 CTD data set).*
- Thompson, J.A.J. Heavy metal analyses of sediments and interstitial water from cores taken in Alice Arm and surrounding environs.*

*General title or description of ongoing study.

APPENDIX F-1

DENSITY AND DISTRIBUTION OF BENTHIC INVERTEBRATES
FOR EACH SAMPLE (0.25 mm screen) FROM ALICE ARM.
ALL RESULTS ARE GIVEN IN NUMBERS PER M².

	CCN		CCM		CCS		DON		DOM		DOS		EEN		EEM		EES		FFM	
	A	B	A	B	A	B	A	B	A	B	A	B	A	B	A	B	A	B	A	B
Coelenterata																				
Hydrozoa																				
Polychaeta																				
Turbellaria																				
Nemertea	10						10				10	10	40				20		160	
Nemertea	240	10			940	1570	4060	640	40	900	170	1780	8260	8320	1450		1180	1870	50480	51080
Annelida																				
Oligochaeta																				
Polychaeta	120		70	50	850	1100	260	870	140		650	840	2230	1320	540		4730	6040	14640	10160
Arthropoda																				
Crustacea																				
Cumacea					10								10		10					
Tenellidae																				
Amphipoda							80	10	10				130	90	70		60	40	120	160
Isopoda							10												40	160
Ostracoda	20		10	20	50	40	250	20	40	60	220	60	710	510	790		60	590	160	120
Haracticulida	10				10	20	40	10	10	30	20	40	10		20		40	20	80	400
Insecta																				
Collembola	10																			
Acarina							10												40	
Mollusca																				
Aplicophora							10													
Gastropoda	770	70			60	30	410	10	120	110	30	100	170	20	280		10	20	160	
Pelecypoda	80	10	20	20	10	10	320	110	50	40		390	280	90	290		510	390	440	280
Echinodermata																				
Ophiuridea																				
Terdigrada			10		10						140	1330	80		40		20	10		40

DENSITY AND DISTRIBUTION OF BENTHIC INVERTEBRATES
FOR EACH STATION (0.25 mm screen) FROM ALICE ARM.
ALL RESULTS ARE GIVEN IN NUMBERS PER M².

	CCN	CCM	CCS	DDN	DDM	DDS	EEN	EEM	EES	FFM
Coelenterata Hydraidea										
Platyhelminthes Turbellaria	5			5		10	20		10	80
Nemertea				2350	470	975	8290	725	1525	50780
Nematoda	125	75	1255							
Annelida Oligochaeta										
Polychaeta	60	60	975	565	70	745	1775	280	5385	12400
Arthropoda Crustacea Cumacea			5				5	5		
Tanaidacea									5	140
Amphipoda				45	5		110	35	50	100
Isopoda				5						20
Ostracoda	10	15	45	135	50	140	610	395	325	140
Harpacticoida	5		15	25	20	30	5	10	30	240
Insecta Collembola	5									
Acarina				5			5			20
Mollusca Aplousobranchia				5						
Gastropoda	420	55	45	210	115	65	95	140	15	80
Pelecypoda	45	20	10	215	45	195	185	145	450	360
Echinodermata Ophiuroidea										
Tardigrada		5	5			735	40	20	15	20

ALL RESULTS ARE GIVEN IN NUMBERS PER M².

	Z2E		Z2M		Z2W		ZSE		Z5M		Z5W	
	A	B	A	B	A	B	A	B	A	B	A	B
Coelenterata Hydroidea	20									50		
Platyhelminthes Turbellaria												
Nemertea	30			10	30		160	10	30	10	180	
Nemertoda	2900		330	680	960	230	2720	1290	1230	4910	1720	310
Annelida Oligochaeta			10									
Polychaeta	10020	8840	5880	5010	4580	2260	13090	11470	9750	410	12100	5720
Arthropoda Crustacea Cumacea		10	70	50	10		180	70	50	200	220	
Tanaidacea			40		10		10	10	20			
Amphipoda		10	20			10		20	10	50		30
Isopoda												
Ostracoda	110	60	10	10	10	10	40	70	20	10	40	30
Harpacticoida	280	120	50	10	170	30	130	60	40	350	200	20
Insecta Collembola												
Acarina	10											
Mollusca Aplousiphora	10							10			10	
Gastropoda	150	70	20	30	30	20	30	20		90	10	10
Pelecypoda	420	390	290	310	230	40	130	140	30	140	40	40
Echinodermata Ophiuroidea												
Tardigrada									10	20	10	20

APPENDIX F-4

DENSITY AND DISTRIBUTION OF BENTHIC INVERTEBRATES
FOR EACH STATION (0.25 mm screen) FROM HASTINGS ARM.
ALL RESULTS ARE GIVEN IN NUMBERS PER M².

	ZZE	ZZM	ZZW	ZSE	ZSM	ZSW
Coelenterata	10				25	
Hydroidea						
Platyhelminthes						
Turbellaria	15	5	15	85	20	90
Nemertea						
Nematoda	1450	505	595	2005	3070	1015
Annelida						
Oligochaeta		5				
Polychaeta	9430	5445	3420	12280	5080	8910
Arthropoda						
Crustacea	5	60	5	125	125	110
Cumacea						
Tanaidacea		20	5	10	10	
Amphipoda	5	10	5	10	30	15
Isopoda						
Ostracoda	85	10	10	55	15	35
Harpacticoida	200	30	100	95	195	110
Insecta						
Collembola	5					
Acarina						
Mollusca						
Aplousobranchia	5			5		5
Gastropoda	110	25	25	25	45	10
Pelecypoda	405	300	135	135	85	40
Echinodermata						
Ophiuroidea						
Tardigrada					15	15

APPENDIX G

LIST OF ALL TAXA FOUND IN ALICE ARM AND HASTINGS ARM

- Porifera
 - Hexactinellida
 - Rosellidae
- Coelenterata
 - Pennatulacea
 - Virgulariidae
 - Virgularia cf. tuberculata Marshal
- Platyhelminthes
 - Turbellaria
 - Polycladida
- Nemertea
 - Anopla
 - Paleonemertea
 - Heteronemertea
 - Lineidae
 - ? Cerebratulus
 - Micrura alaskensis? Coe
- Enopla
 - Haplonemertea
- Nematoda
- Sipunculida
 - Golfingiidae
 - Golfingia sp.
- Echiura
- Annelida
 - Oligochaeta
 - Tubificida
 - Tubificidae
 - Limnodriloides sp.
 - L. victoriensis Brinkhurst and Baker
- Polychaeta
 - (Errantiate)
 - Polynoidae
 - ? Eunoe
 - Gattyana treadwelli? Pettibone

Sigalionidae

Pholoe minuta (Fabricius)

Phyllodocidae

Eteone (Mysta) sp.

E. columbiensis Kravitz and Jones

Phyllodoce groenlandica Oersted

Pilargidae

Ancistrostylis sp.

A. groenlandica McIntosh

Syllidae

Exogone sp. 1

Exogone sp. 2

E. gemmifera Pagenstecher

Eusyllis cf. blomstrandi Malmgren

Syllis sp.

S. alternata Moore

Nephtyidae

Aglaophamus malmgreni (Theel)

Nephtys cornuta cornuta Berkley and Berkley

N. punctata Hartman

Sphaerodoridae

Sphaerodoropsis sphaerulifer (Moore)

Goniadidae

Goniada annulata Moore

Glycinde armigera Moore

Lumbrineridae

Lumbrineris sp.

L. luti Berkley and Berkley

Paraninoe simpla (Moore)

Dorvilleidae

Schistomeringos sp.

Hesionidae

Gyptis brevipalpa (Hartmann-Schroder)

(Sedentariate)

Orbiniidae

Leitoscoloplos pugettensis (Pettibone)

Paraonidae

Aricidea suecica Eliason

A. lopezi lopezi Berkley and Berkley

Cirrophorus branchiatus Ehlers

Levisenia gracilis (Tauber)

Trochochaetidae

Trochochaeta multisetosa (Oersted)

Chaetopteridae

Spiochaetopterus costarum (Claparede)

Spionidae

Polydora sp.

Pseudopolydora kemp? (Southern)

Prionospio sp.

P. cirrifera Wiren

P. steenstrupi Malmgren

Spiophanes sp.

S. kroyeri Grube

Cirratulidae

Chaetozone setosa Malmgren

Caulleriella hamata (Hartman)

C. cf. hamata (Hartman)

Cossuridae

Cossura soyeri Laubier

Flabelligeridae

Brada sp.

B. villosa (Rathke)

Pherusa sp.

Opheliidae

Ophelina breviata (Ehlers)

Sternaspidae

Sternaspis scutata (Renier)

Capitellidae

Capitella capitata (Fabricius)

Decamastus sp.

Heteromastus sp.

H. filobranchus? Berkley and Berkley

Mediomastus sp.

Maldanidae

Maldane glebifex Grube

Rhodine sp.

Oweniidae

Myriochele oculata Zachs

Owenia fusiformis Chiaje

Amphictenidae

Pectinaria sp.

P. moorei Annenkova

Terebellidae

Artacama conifera Moore

Pista cristata (Muller)

Polycirrus sp.

Trichobranchidae

Terebellides sp.

Terebellides stroemi Sars

Ampharetidae

Amage anops (Johnson)

Amphicteis sp.

A. cf. scaphobranchiata Moore

Anobothrus gracilis (Malmgren)

Sosanopsis cf. hesslei Banse

Sabellidae

Euchone sp.

Jasmineira pacifica Annenkova

UID Polychaeta

Arthropoda

Crustacea

Cumacea

Campylaspis ? papillata Lomakina

Eudorella sp.

Leucon sp.

Tanaidacea

Leptognathia sp.

Amphipoda

Corophiidae

Corophium acherusicum Costa

Lysianassidae

Cyphocaris challenger Stebbing

Koroga megalops Holmes

Orchomene obtusa (Sars)

Pachynus barnardi Hurley

Phoxocephalidae

Paraphoxus oculatus (Sars)

Oedicerotidae

Bathymedon pumilis Barnard

Monoculodes cf. emarginatus Barnard

Eusiridae

?Rhacotropis sp.

Hyperiidæ

Hyperia sedusarum Muller

Parathemisto sp.

P. pacificus Stebbing

Mysidacea

Mysidæ

Pseudomma truncatum S.I. Smith

Decapoda

Mollusca

Aplacophora

Gastropoda

Pyramidellida

Pyramidellidæ

Turbonilla sp.

Cephalospidea

Cylichna altonsa

Gastropteron pacificum? Bergh

Thecosomata

Limacina sp.

Neogastropoda

Thaididæ

Buccinidæ

Buccinum sp.

Megogastropoda

Vitrinellidæ

?Vitrinella sp.

Scaphopoda

Dentaloida

Dentaliidæ

Dentalium rectius Carpenter

Siphonodentaloida

Cadulidæ

Cadulus californicus (Pilsbury and Sharp)

Bivalvia

Heterodonta

Tellinidæ

Macoma sp.

M. carlottensis Whiteaves

M. elimata Dunnill and Coan

M. cf. nasuta (Conrad)

Cooperellidae
Cooperella sp.

Veneridae
Transenella tantilla (Gould)

Ungulidae
Diplodonta orbella Gould

Hiatellidae
Hiatella arctica (Linnaeus)
Nuculana minuta Fabricius
Nucula tenuis (Montagu)
Yoldia amygdalea Valenciennes
Y. beringiana Dall
Y. martyria Dall
Y. myalis (Couthouy)

Echinodermata
Asteroidea
Paxilloridea
Goniopectinidae
Ctenodiscus crispatus Retzius

Ophiuroidea
Ophiurida
Ophiolepididae
Ophiura sp.
O. leptoctenia H.L. Clark
O. sarsi Lutken

Holothuroidea
Apodida
Chirodotidae
Chirodota albatrossi Ohshima

Molpadida
Molpadiidae
Molpadia intermedia (Ludwig)

Hemichordata
Enteropneusta

DENSITY AND DISTRIBUTION FOR BENTHIC INVERTEBRATES (1.0 mm screen) FROM ALICE ARM.
ALL RESULTS ARE GIVEN IN NUMBERS PER M².

	CCN	CCM	CCS	DDN	DDM	DOS	EEN	EEM	EES	FFM
	A B	A B	A B	A B	A B	A B	A B	A	A B	A B
Pollera Hexactinellida Rosettidae							10 S			
Cactenierata Pennatulacea Virgulariidae <i>Virgularia cf. Nidocaulis</i>										
Platyhelminthes Turbellaria Polychaeta									10 20 15	60 30
Nemertea Annelida Polemonerites										
Meteromerites Linnidae <i>? Ceratophyllus</i> <i>Microstomum</i> <i>Microstomus</i>							10 S		90 50 70 10 5 10 5	
Enopla Haplomerites				10 S						
Nematoda							10 S		10 S	240 270 255
Sipunculida Gastropoda <i>Gastropoda sp.</i>										
Echinozoa										
Annelida Oligochaeta Tubificidae Tubificidae <i>Lumbricoides sp.</i> <i>L. terrestris</i>										10 10 10 20 30 25
Polychaeta (Errantia) Polychaeta <i>? Eurydora</i> <i>Gastropoda</i>										10 10 10 S S
<i>Gastropoda</i>			20 10				10 S			90 50 70

[illegible]

[illegible]

[illegible]

[illegible]

DENSITY AND DISTRIBUTION FOR BENTHIC INVERTEBRATES (1.0 mm screen) FROM HASTINGS ARM.
ALL RESULTS ARE GIVEN IN NUMBERS PER M².

	ZZE	ZZM	ZZW	ZSE	ZSM	ZSW
A B X	A B X	A B X	A B X	A B X	A B X	A B X
Parifera Hexactinellida Rosellidae						
Coelenterata Pennatulacea Virgulariidae <u>Virgularia cf. tuberculata</u>	20 10					10 5
Platyhelminthes Turbellaria Polychaetida			10 5			
Nemertea Anopla Paleonemertea	10 5	20 10	60 20 40	10 20 15	20 30 25	30 15
Heteronemertea Lineidae ? <u>Cerebratulus</u> <u>Micrura alaskensis?</u>	10 5	10 5		10 5	20 10	20 10
Ecnepha Haploneurtemea			10 5			
Nematoda						
Sipunculida Golfingiidae <u>Golfingia sp.</u>						
Echiura						
Annelida Oligochaeta Tubificidae Tubificidae <u>Limnodriloides sp.</u> <u>L. victorialis</u>						
Polychaeta (Errantiate) Polynoidae ? Eunoe <u>Gattyana treadwellii?</u>	10 5	10 5		20 10 10	10 5	

	ZSE			ZSM			ZSE			ZSM			ZSW		
	A	B	R	A	B	R	A	B	R	A	B	R	A	B	R
Sigallanidae <u>Phaloe minuta</u>	10		5												
Phyllodocidae <u>Eteone (Myrta) sp.</u> <u>E. columbiensis</u> <u>Phyllodoce groenlandica</u>															
Pillargidae <u>Ancistrayllis sp.</u> <u>A. groenlandica</u>	10		5												
Syllidae <u>Exogone sp. 1</u> <u>Exogone sp. 2</u> <u>E. gemmifera</u> <u>Euryllis cf. blomstrandii</u> <u>Syllis sp.</u> <u>S. alternata</u>	30 570	30 200	30 385	10 180	200 20	5 190 20	20 520	30 190	25 355	20 520	30 190	5 355	10 420	5 400	5 350
Nephtyidae <u>Aglaophanus malmgreni</u> <u>Nephtys cornuta cornuta</u> <u>N. punctata</u>															
Sphaerodoridae <u>Sphaerodoropsis sphaerulifer</u>															
Coniadiidae <u>Coniada annulata</u> <u>Glycinde armigera</u>	10	30	20	20	20	20	20	40	30	20	40	30	40	30	15
Lumbrineridae <u>Lumbrineris sp.</u> <u>L. luti</u> <u>Paraninos simplex</u>	20	10	15		10	5	10	10	5		20	10	30	20	25
Darvillidae <u>Schiostomeringos sp.</u>															
Hesionidae <u>Cyrtis brevicauda</u>	30		15	20	50	35	50	30	40	20	70	45	100	80	105
(Sedentariidae) Orbinidae <u>Leitocarpus pugettensis</u>							10		5				10		5

[illegible]

	ZZE	Z2M	Z2W	ZSE	ZSM	ZSW
A B X	A B X	A B X	A B X	A B X	A B X	A B X
Oveniidæ <u>Myriochela acutata</u> <u>Ovenia fusiformis</u>				10 10 10	10 10 10	10 10 5
Amphictenidæ <u>Pectinaria sp.</u> <u>P. moorei</u>	30 15 5	10 10 5		10	10 10 10	10 10 5
Terebellidæ <u>Artacama conifera</u> <u>Pista cristata</u> <u>Polysira sp.</u>				10	10 10 10	
Trachobranchiidæ Terebellidæ sp. <u>T. stromi</u>	10 5					
Ampharetidæ <u>Anagapopsis</u> <u>Amphicteta sp.</u> <u>A. cf. scaphobranchiata</u> <u>Anabittus gracilis</u> <u>Scolecops cf. heslei</u>		10 10 5				
Sabellidæ <u>Euchone sp.</u> <u>Laminella pacifica</u>						
UD Polychœta						
Arthropoda						
Crustacea						
Cumacea <u>Comptaspis ? papillata</u> <u>Eudrella sp.</u> <u>Leucon sp.</u>	10 5	50 50 50	30 15	100 90 95	70 50 60	80 20 50
Tanaidacea <u>Leptognathia sp.</u>			10 5			10 5
Amphipoda Corophiidæ <u>Corophium apertum-like</u>						
Lydianellidæ <u>Cyphocaris chailangeni</u> <u>Karaga megalyra</u> <u>Psidium polinae</u> <u>Psidium parvum</u>					10	10 5
Phoronopodidæ <u>Paragonomus setulosus</u>						

	ZZE	Z2M	Z2W	ZSE	ZSM	ZSW
A B X̄	A B X̄	A B X̄	A B X̄	A B X̄	A B X̄	A B X̄
Oedicerotidae <u>Bathymedon pumilis</u> <u>Monaculodes cf. emarginatus</u>	20 10	10 5		20 10	10 10 5	10 10 10 5
Eusiridae ? <u>Rhodotropis sp.</u>						
Hyperiliidae <u>Hyperilia sedusarum</u> <u>Parathemista sp.</u> <u>P. pacificus</u>			10 5			10 5
Mysidacea Mysidae <u>Pseudomima truncatum</u>						
Decapoda						
Mollusca Aplousobranchia						10 5
Gastropoda Pyramidelidae Pyramidellidae <u>Turbanilla sp.</u>						10 5
Cephalopoda <u>Cylindrina allansea</u> <u>Gastropheron pacificum?</u>	10 5	40 50 45	20 60 40	20 70 45	50 100 75	70 30 50
Thecosomata <u>Limnoria sp.</u>					10 5	
Neogastropoda Thaididae						
Buccinidae <u>Buccinum sp.</u>						
Megagastropoda Vitrinellidae <u>?Vitrinella</u>						
Scaphopoda Dentaliida Dentaliidae <u>Dentalium rectius</u>	10 5	10 5		10 5	10 20 15	20 10 15
Siphonodentaliida Caduliidae <u>Cadulus californicus</u>	10 5				10 5	

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ADDENDUM

After the manuscript was in press, a number of incorrect or outdated familial or generic assignments were discovered. The following classification and specific names, recognized by Austin (1983), should replace those presented in Appendices G, H, and I.

- Porifera
 - Hexactinellida
 - Lyssacinosa
 - Rosellidae
- Coelenterata
 - Anthozoa
 - Pennatulacea
 - Virgulariidae
 - Virgularia cf. tuberculata Marshal
- Platyhelminthes
 - Turbellaria
 - Polycladida
- Nemertea
 - Anopla
 - Palaeonemertea
 - Heteronemertea
 - Lineidae
 - ? Cerebratulus
 - Micrura alaskensis? Coe
- Enopla
 - Hoploneurtea
- Nematoda
- Sipunculoidea
 - Golfingiidae
 - Golfingia sp.
- Echiuroidea
- Annelida
 - Oligochaeta
 - Tubificida
 - Tubificidae
 - Limnodriloides sp.
 - L. victoriensis Brinkhurst and Baker

Paraonidae

Aricidea suecica Eliason

A. lopezi lopezi Berkley and Berkley

Cirrophorus branchiatus Ehlers

Levinsenia gracilis (Tauber)

Trochochaetidae

Trochochaeta multisetosa (Oersted)

Chaetopteridae

Spiochaetopterus costarum (Claparede)

Spionidae

Polydora sp.

Pseudopolydora kempi? (Southern)

Prionospio sp.

P. cirrifera Wiren

P. steenstrupi Malmgren

Spiophanes sp.

S. kroyeri Grube

Cirratulidae

Chaetozone setosa Malmgren

Cauleriella hamata (Hartman)

C. cf. hamata (Hartman)

Cossuridae

Cossura soyeri Laubier

Flabelligeridae

Brada sp.

B. villosa (Rathke)

Pherusa sp.

Opheliidae

Ophelina breviata (Ehlers)

Sternaspidae

Sternaspis scutata (Renier)

Capitellidae

Capitella capitata (Fabricius)

Decamastus sp.

Heteromastus sp.

H. filobranchus? Berkley and Berkley

Mediomastus sp.

Maldanidae

Maldane glebifex Grube

Rhodine sp.

Polychaeta
(Errantiate)

Polynoidae

? Eunoe

Gattyana treadwelli? Pettibone

Sigalionidae

Pholoe minuta (Fabricius)

Phyllodocidae

Eteone (Mysta) sp.

E. columbiensis Kravitz and Jones

Phyllodoce groenlandica Oersted

Pilargidae

Ancistrosyllis sp.

A. groenlandica McIntosh

Syllidae

Exogone sp. 1

Exogone sp. 2

E. gemmifera Pagenstecher

Eusyllis cf. blomstrandii Malmgren

Syllis sp.

S. alternata Moore

Nephtyidae

Aglaophamus malmgreni (Theel)

Nephtys cornuta cornuta Berkley and Berkley

N. punctata Hartman

Sphaerodoridae

Sphaerodoropsis sphaerulifer (Moore)

Goniadidae

Goniada annulata Moore

Glycinde armigera Moore

Lumbrineridae

Lumbrineris sp.

L. luti Berkley and Berkley

Paraninoe simpla (Moore)

Dorvilleidae

Schistomeringos sp.

Hesionidae

Gyptis brevipalpa (Hartmann-Schroder)

(Sedentariate)

Orbiniidae

Leitoscoloplos pugettensis (Pettibone)

Oweniidae

Myriochele oculata Zachs

Owenia fusiformis Chiaje

Amphictenidae

Pectinaria sp.

P. moorei Annenkova

Terebellidae

Artacama conifera Moore

Pista cristata (Muller)

Polycirrus sp.

Trichobranchidae

Terebellides sp.

Terebellides stroemi Sars

Ampharetidae

Amage anops (Johnson)

Amphicteis sp.

A. cf. scaphobranchiata Moore

Anobothrus gracilis (Malmgren)

Sosanopsis cf. hesslei Banse

Sabellidae

Euchone sp.

Jasmineira pacifica Annenkova

UID Polychaeta

Arthropoda

Crustacea

Cumacea

Nannastacidae

Campylaspis ?papillata Lomakina

Leuconidae

Eudorella sp.

Leucon sp.

Tanaidacea

Leptognathiidae

Leptognathia sp.

Amphipoda

Corophiidae

Corophium acherusicum Costa

Lysianassidae

Cyclocaris challenger Stebbing

Koroga megalops Holmes

Orchomene obtusa (Sars)

Pachynus barnardi Hurley

Phoxocephalidae

Paraphoxus oculatus Sars

Oedicerotidae

Bathymedon pumilis Barnard

Monoculodes cf. emarginatus Barnard

Eusiridae

?Rhachotropis sp.

Hyperiididae

Hyperia medusarum (Muller)

Parathemisto sp.

P. pacifica Stebbing

Mysidacea

Mysidae

Pseudomma truncatum S.I. Smith

Decapoda

Mollusca

Aplacophora

Gastropoda

Pyramidellida

Pyramidellidae

Turbonilla sp.

Cephalaspidea

Cylichnidae

Cylichna altonsa

Gastropteridae

Gastropteron pacificum? Bergh

Thecosomata

Limacinidae

Limacina sp.

Neogastropoda

Nucellidae

Buccinidae

Buccinum sp.

Mesogastropoda
Vitrinellidae
? Vitrinella sp.

Scaphopoda
Dentaliida
Dentaliidae
Rhabdus rectius Carpenter

Gadilida
Siphonodentaliidae
Polyschides californicus (Pilsbry and Sharp)

Bivalvia
Veneroida
Tellinidae
Macoma sp.
M. carlottensis Whiteaves
M. eliminata Dunnill and Coan
M. cf. nasuta (Conrad)

Cooperellidae
Cooperella sp.

Veneridae
Transenella tantilla (Gould)

Ungulinidae
Diplodonta orbella Gould

Hiatellidae
Hiatella arctica (Linnaeus)

Nuculoidae
Nuculanidae
Nuculana minuta Fabricius

Nuculidae
Nucula tenuis (Montagu)

Yoldiidae
Yoldia amygdalea Valenciennes
Y. beringiana Dall
Y. martyria Dall
Y. myalis (Couthouy)

Echinodermata
Asteroidea
Paxillosida
Goniopectinidae
Ctenodiscus crispatus Retzius

Ophiuroidea
Ophiurida
Ophiuridae
Ophiura sp.
O. leptoctenia H.L. Clark
O. sarsi Lutken

Holothuroidea
Apodida
Chiridotidae
Chiridota albatrossi Ohshima

Molpadida
Molpadiidae
Molpadia intermedia (Ludwig)

Hemichordata
Enteropneusta

Austin, W.C. 1983. An annotated checklist of marine invertebrates in the cold temperate Northeast Pacific. Khoyatan Marine Laboratory, Cowichan Bay, B.C. 621 pp.

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